# The Hyperviscosity Syndromes

# HAU C. KWAAN, M.D., PH.D. AND ANUREKHA BONGU, M.D.

ABSTRACT Impaired blood flow due to abnormal rheologic characteristics results in a multiplicity of clinical manifestations, collectively termed the hyperviscosity syndrome. A basic knowledge of the principles of rheology is important in the understanding of its pathophysiology, especially the relationship between viscosity and flow conditions. The flow characteristics in different types of blood vessels are also determinants in the location of the clinical manifestation. The syndrome can occur in a wide variety of diseases and is best grouped according to the causative element or elements in blood. Abnormalities in the cellular components of blood can occur in the quantity and the quality of erythrocytes, leukocytes, and platelets. Abnormal plasma components can also be in both the quantity and quality of the plasma proteins. Clinical manifestations are the result of vascular occlusion, especially in the microcirculation. The altered rheologic characteristics of either the cellular or the protein component may be temperature dependent, being abnormal only at temperatures below 37°C, so that only the cooler parts of the body are affected. The management of these conditions should be primarily directed at the removal of the abnormal component. At the same time, it should be accompanied by measures that can control the production of the causative element.

Keywords: Hyperviscosity, hemorheology, cancer, thrombosis, hemostasis

The hyperviscosity syndromes consist of a group of pathologic conditions in which blood flow is impaired due to abnormal flow characteristics of the blood. Many thromboembolic complications of hyperviscosity occur

Division of Hematology/Oncology, Department of Medicine, Northwestern University Medical School, Chicago, Illinois.

Reprint requests: Dr. Kwaan, Division of Hematology/Oncology, Lakeside Division, VA Healthcare System, 333 East Huron Street, Chicago, IL 60611-3004.

in the setting of underlying neoplastic or paraneoplastic conditions, and are thus of important consideration for discussion in the current issue devoted to the thrombophilic state in cancer. To recognize and correctly diagnose the clinical manifestations of hyperviscosity, the clinician needs to have a clear understanding of its pathophysiology and its relevance to the commonly associated diseases in order to provide the proper management.

## RHEOLOGIC CONSIDERATIONS

The flow properties of blood are dependent on both its fluid (plasma) and cellular contents. The hemodynamic factors influencing the flow of blood through blood vessels have been well defined and the reader may wish to refer to detailed descriptions in a number of reviews.1-4 Certain commonly used rheologic terms need to be first defined. The velocity of flow through a cylindrical vessel is a function of the radial distance from the vessel wall, being highest in the center line of flow (axial flow) and lowest adjacent to the vessel wall (marginal flow). The relative velocity of flow through parallel fluid layers is termed the shear rate, being a measurement of how fast adjacent planes of fluid slide past each other. Fluid at the margin of a vessel flows at a higher shear rate than that occurring in axial (central) flow. Viscosity is a measure of the resistance to deformation of a fluid. The force exerted on the vessel wall during the flow is termed wall shear stress, and is a function of the shear rate and the viscosity of the blood. The flow rate, hence the shear rate, is much higher in arterioles than in veins.

Thus, provided that the viscosity remains unchanged, the wall shear stress is also much higher in arterioles than in veins, and can be quite high in stenotic vessels. A high shear stress has functional significance in that it increases platelet aggregation.5,6 This form of modulation of endothelial cell metabolism plays an important role in atherogenesis and increased thrombotic risk in stenotic vessels where the shear stress is high. The viscosity of whole blood is a function of the plasma viscosity, hematocrit, and red cell aggregation. In a fluid

Copyright © 1999 by Thieme Medical Publishers, Inc., 333 Seventh Avenue, New York, NY 10001. Tel: +1 (212) 760-0888, ext. 132. 0094-6176/1999/E1098-9064(1999)25:02:0199-0208:STH00512X

suspension of noncompressible particles (known as a Newtonian fluid), the shear stress is a linear function of the shear rate. In contrast, whole blood is a nonNewtonian fluid, in which the shear stress increases greatly with any increase in the viscosity of blood, especially at low shear rates. Whole blood viscosity is dependent on both the cellular and the plasma contents of blood. A high hematocrit has a higher viscosity at all shear rates. Factors governing the plasma viscosity are the concentration of plasma proteins and the characteristics of the proteins. For example, the viscosity of blood is directly proportional to the plasma fibrinogen level. This was well demonstrated by Bell et al<sup>7</sup> in therapeutic defibrination of patients with Ancrod (Fig. 1). At a normal fibrinogen level, the blood is a Newtonian fluid, whereas after defibrination, at fibrinogen levels of 5 mg/dL, the viscosity is not only decreased but is a direct function of the shear rate, behaving as a Newtonian fluid. Increased gammaglobulin levels and their abnormal characteristics (such as large molecular size in macroglobulinemia) are also major determinants in hyperviscosity syndromes in plasma cell dyscrasias. Flow is also affected in conditions where abnormally increased red cell aggregation occurs, especially in the microcirculation. At low shear rates, red cell aggregation leads to a greater viscosity, while at high shear rates, red cell aggregates are deformed (disaggregated), resulting in a lower viscosity8 (Fig. 2). Abnormal deformability of red cells, as seen in sickle cell disease,9 influences the nature of the red cell aggregates and hence whole blood viscosity.

# EFFECTS OF SHEAR STRESS AND CYCLICAL STRAIN ON ENDOTHELIAL AND SMOOTH MUSCLE CELLS

In addition to shear stress, the vascular wall is also subjected to cyclical strain caused by the pulsatile circumferential stress from the pulsatile pump action of the heart, more so on the arterial than on the venous wall.<sup>10</sup> An important biologic role of both shear stress and cyclic strain is their modulation of the expression of proteins by both endothelial and smooth muscle cells in the vascular wall. These proteins are involved in a wide range of cellular functions.1 Some of these need special consideration. Endothelin-1 is a peptide produced by endothelial cells that is both a vasoconstrictor and smooth muscle cell mitogen. While increased shear stress results in a decrease in endothelin-1, cyclical strain increases its synthesis and secretion.11 The plasminogen/plasmin system is also affected. A high shear stress increases the endothelial synthesis and secretion of t-PA but not PAI-1, whereas cyclical strain increases PAI-1 without affecting t-PA.<sup>12,13</sup> Endothelial synthesis of prostacyclin, a potent vasodilator and antiplatelet aggregating peptide, is increased by increased shear stress<sup>14</sup> and cyclical strain.<sup>15</sup> These changes in protein synthesis are believed to be

due to stress modulation of the respective gene expression and activation of multiple transcription factors such as AP-1, NFkB, Sp-1, and Egr1.<sup>16,17</sup> Though much of these studies are aimed at the pathogenesis of thrombosis and arteriosclerosis, the findings can also be applied to tumor biology. The influence of the altered blood flow in tumors and in metastatic lesions has largely been ignored until recently, and should well be a fertile area of future investigations.

#### CELLULAR COMPONENTS OF BLOOD

The viscosity of blood is dependent on the concentration of its cellular components or its cytocrit. A highpacked cell volume can be caused by an increase in anyone of the cellular components, red cells, leukocytes, or platelets. An increase in blood viscosity can result in anyone of these conditions.

#### **Increased Hematocrit**

Blood viscosity is raised in commonly encountered clinical situations in which the hematocrit is increased from a shrunken plasma volume, such as in dehydration and in spurious polycythemia (Gaisbock syndrome). One of the most common causes of a high hematocrit in cancer patients is dehydration. However, its significance in causing an increased viscosity is not always recognized. Loss of water in the plasma compartment may also increase the protein concentration and this, in turn, raises the viscosity through a number of ways, including an increase in red cell aggregation. A high hematocrit also results in an increase in the axial migration of the red cells while the marginal plasmatic zone narrows. This will result in an increase in the shear rate and shear stress and will allow a greater platelet-endothelial cell interaction. At higher shear rates, the binding of von Willebrand factor to GpIb and to GpIIb/IIa on platelet cell surface is increased, and thus resulting in platelet activation.6,18

The various causes of erythrocytosis with a high hematocrit are listed in Table 1. The most common cause of erythrocytosis that gives serious concerns for thromboembolic complications is polycythemia vera. In this condition, an increased risk for thromboembolic complications has been long recognized. The data derived from the Polycythemia Study Group indicated that over 10 years of the study, thrombosis was the most common cause of mortality accounting for 29% of the deaths.<sup>19</sup> These data are strongly supported by an Italian study of 1213 patients over 20 years,<sup>20</sup> showing that thromboembolic complications account for 30% of the deaths. Among the initial thrombotic events, cerebral vascular accidents were the most frequent (34%), followed by myocardial infarction (13%), and peripheral arterial occlusion (9%), while venous thrombosis accounted for



**FIG. 1.** Blood viscosity following defibrinating therapy with Ancrod, showing the rheologic properties of the blood change as the fibrinogen level falls with the therapy. The normal nonNewtonian characteristics of normal blood change to those of a Newtonian fluid as the fibrinogen is removed progressively. (Reprinted with permission from reference 7.)

26% of these events.<sup>19,20</sup> Of significance is the finding that 35% of the thrombotic events were fatal. The high rate of thrombotic events in the cerebral circulation has been found to be due to impaired cerebral blood flow as the result of increased arterial oxygen. The cerebral



TABLE 1. Conditions Associated with Erythrocytosis.

Increased erythropoietin production (kidney neoplasms, polycystic kidneys, hepatocellular carcinoma, cerebellar hemangioblastoma, uterine leiomyoma, adrenal neoplasms, drug-induced; by erythropoietin or androgens)

blood flow returns to normal on reduction of the hematocrit.

The management of polycythemia vera is well described in many reviews<sup>21–23</sup> and will not be detailed here. The main objective is to reduce the whole blood viscosity to prevent thromboembolic complications. Increase in whole blood viscosity is proportional to the



**FIG. 2.** Logarithmic relation between relative apparent viscosity ( $\eta_r$ ) and apparent shear rate ( $\gamma_a$ ) for suspensions of human normal red cells in autologous plasma and in albumin-Ringer solution. In both suspensions, the cell volume concentration is 45% and the external fluid viscosity is 1.2 cP<sup>-</sup> with the temperature at 37°C. Drawings show the aggregation of red cells in plasma, but not in albumin-Ringer solution, at low shear rates. Note the deformation into monodispersed red cells in both suspensions at high shear rates. Reprinted with permission from reference 8.

shear rate of blood flow. At a moderately raised hematocrit level of 60%, for example, the whole blood viscosity is twice that of blood with a hematocrit of 40% at shear rates of 12 per sec, calculated to be the flow conditions in a medium arteriole.<sup>24</sup> Thus, the mainstay of therapy is the reduction of the hematocrit. This should be carried out by the least toxic and leukemogenic measures for the whole duration of the disease.

#### Abnormal Red Cell Aggregation

In addition to the increased viscosity caused by an increase in the hematocrit, blood flow may be impaired by abnormal rheologic properties of the red cells. The most frequent is an increased red cell aggregation. Different forms of aggregation can occur, the best known being rouleaux formation observed in the peripheral blood smears of patients with plasma cell dyscrasias. Detailed in vitro studies by Dintenfass and others have revealed that in normal blood, red cell aggregates of six to 20 cells may occur in experimental flow models.<sup>25,26</sup> These small aggregates break down easily and do not cause a block in the microcirculation. In contrast, large aggregates were observed in diabetes, hyperlipidemia, and certain forms of cancer, with each single aggregate consisting of up to 10<sup>5</sup> red cells.<sup>27,28</sup> These large aggregates do not disperse easily and are believed to be the cause of impaired microcirculation. We have observed abnormally intense red cell aggregation in a number of patients with acquired dysfibrinogenemia of unknown etiology (Fig. 3A, B).<sup>29</sup> These patients had symptoms of microvascular occlusion. In one patient, following therapeutic defibrination with Ancrod, the red cell aggregation decreased with a dramatic relief of the ischemic symptoms.<sup>29</sup> We have also used low molecular weight Dextran (Rheomacrodex) to treat patients in whom we had demonstrated the presence of abnormally increased red cell aggregation caused by their abnormal fibrinogen. Invariably the peripheral microvascular occlusive manifestations abated following an improvement in objective observations of decreased red cell aggregation (unpublished observations). Interestingly, experiments performed on the space shuttle orbiter "Discovery" showed that the abnormal red cell aggregation found at ground level reverted to normal red cell distribution at zero gravity, suggesting that gravity may play a role in the cell–cell interaction.<sup>30</sup>

#### Increased Leukocrit

Impaired microcirculation can result from either a high leukocyte count with a greatly increased leukocrit or abnormal leukocytes in acute leukemia. Under normal circumstances, leukocytes respond to inflammatory stimuli by margination and extravasation from the circulating blood. This process consists of several steps beginning with contact of the leukocytes in the marginal zones of the vessel wall, followed by rolling of the cells along the endothelial surface, then by firm adhesion to the endothelium, and, finally, by diapedesis or migration through the wall. This series of steps is modulated by a number of cell adhesion molecules (CAMS).31-33 The CAMS are classified into four families: (a) The Ig superfamily of CAMS, which are cell surface glycoproteins expressed on endothelial cells, consisting of ICAM-1 (CD54), ICAM-2 (CD102), ICAM-3 (CD50), and VCAM-1 (CD106), and on platelets of PECAM (CD31). By binding to other adhesive molecules, they enhance the adhesion between the endothelium and leukocytes.34,35 (b) The integrins, which are heterodimeric transmembrane glycoproteins involved in cell-cell adhesion and cell-matrix interactions.36,37 A good example is  $\alpha_{\text{ub}}\beta_3$  (GpIIb/IIIa or CD41/61) on the platelet surface, which on binding with von Willebrand factor initiates platelet adhesion to vessel wall and subsequent aggregation. In leukocytes, the integrin  $\alpha_1 \beta_2$  (CD11a/18)(or



FIG. 3. Ischemic changes in the digits of a patient with increased red cell aggregation due to an acquired dysfibrinogenemia showing gangrene of two fingers in the left hand (A). A cover-slip preparation of his blood shows marked *rouleaux* formation of his red cells in long chains (B). (Reprinted with permission from reference 29.)

LFA-1) in lymphocytes, the integrin  $\alpha_M \beta_2$  (CD11b/ 18)(or Mac-1) and the integrin  $\alpha_{x}\beta_{2}$  (CD11c/18)(or p150/95) in neutrophils and monocytes are responsible for binding of these respective leukocytes to the endothelial surface and are inducible by inflammatory cytokines.38,39 (c) The cadherins are also transmembrane glycoproteins and mediate cell adhesion through a calcium-dependent mechanism.40,41 They are believed to be involved in tumor invasion and metastasis.42 (d) The selectin family, consisting of E-selectin (CD62E) in endothelial cells, P-selectin (CD62P) in both platelets and endothelial cells and L-selectin (CD62L) in leukocytes.43-45 The selectins recognize ligands with carbohydrate structures such as a 150 kDa protein E-selectin ligand-1, expressed in leukocytes. Another example is the ligand sialyl-Lewis X, expressed in cancer cell surfaces, which mediates the adhesion of tumor cells to endothelium through E-selectin.46

In leukostasis, these adhesive molecules are believed to be taking part in the adhesion and extravasation of leukocytes, though this process has not been extensively studied. They have also been shown to be involved in the metastatic spread of tumor cells.47-49 Leukostasis is encountered when the whole blood viscosity is increased from either a very high leukocyte count, as in chronic leukemias, or from a high myeloblast or lymphoblast count in acute leukemias, and in high-grade nonHodgkin's lymphoma. Whole blood viscosity is increased when a leukocrit of 20 to 25% is reached, as seen with 4 to  $6 \times 10^5$  myeloblasts/µL or 5 to  $10 \times 10^5$  lymphoblasts or lymphocytes/µL. The risk of this complication is highest with acute myeloblastic leukemia because the myeloblasts are less deformable. Because the process of leukocyte/endothelial cell interaction can be activated by inflammatory cytokines, such as IL-1b and TNF $\alpha$ , a local inflammatory process can often precipitate leukostasis of leukemic cells. Leukostasis under local inflammatory conditions can thus occur at a lower leukocyte count. An example is the occurrence of pulmonary leukostasis in a leukemic patient with pneumomia.<sup>50</sup> The properties of the activated leukocytes are altered by the cytokines allowing a greater adhesion to the vessel wall. For example, the activated leukocyte is flattened out on the endothelial surface, allowing increased bonding to the endothelium. Additionally, the altered hemodynamics of decreased fluid drag and torque on the flattened leukocytes results in greater adhesion.51 There is evidence that the E-selectin is activated in Hodgkin's disease and anaplastic large cell lymphoma.52 In the lymph nodes of these diseases, a good correlation was found between a high E-selectin expression in the lymphoma cells and the presence of intravascular thrombi and the immuno-histochemical identification of tissue factor in the endothelial cells. In contrast, the Eselectin expression and tissue factor staining in nonmalignant reactive lymph nodes are uncommon. The resulting occluded microcirculation is due to both adhesion

and aggregation of abnormal leukocytes. Due to the local anatomical characteristics of blood vessels in the brain and the lungs, the leukostasis syndrome manifests most commonly in these organs, although histologic evidence of leukostasis can be also found in other parts of the body, especially in the liver and the spleen.53 Postmortem studies by McGee et al54 have shown that all patients with leukocyte counts greater than 2  $\times$  10<sup>5</sup> cells/µL and 50% of those with leukocyte counts between  $0.5 \times 10^5$  cells/µL to  $2 \times 10^5$  cells/µL had prominent leukocyte aggregates in their tissues. Autopsy findings of the brain indicate that central nervous system leukostasis is more intensive in the medium-sized vessels in the white matter and the leptomeninges, and surprisingly affecting less frequently the cortical capillaries.55 Leukocyte aggregates are present in the occluded vessels, and, in the case of acute leukemia, vessel wall penetration with perivascular infiltration ("cuffing") by myeloblasts or lymphoblasts can often be seen. Clinical manifestations of leukostasis in the brain are symptoms that range from headache to coma and death, with signs of increased intracranial pressure, often accompanied by focal neurologic deficits.56 Leukostasis in the pulmonary circulation presents with acute respiratory failure and with bilateral pulmonary infiltrates prominent in the chest X ray.57, 58

Leukostasis, in patients that are symptomatic, require emergency treatment with leukapheresis. Large number of cells in the range of 10<sup>11</sup> to 10<sup>12</sup> cells can be removed in a single apheresis procedure,<sup>59,60</sup> depending on the patient's leukocyte count. Due to the smaller volume of blood removed in leukapheresis, most patients can tolerate the procedure better than plasma exchange. Prompt initiation of hydroxyurea administration can lower the white count, and thus the leukocrit, to bring about a decrease in the whole blood viscosity.<sup>61,62</sup> For patients that are asymptomatic, this form of treatment may suffice without using leukapheresis. Concurrent chemotherapy for the leukemia should also be given.

#### Thrombocytosis

A high platelet count has been found to be associated with an increased risk of thrombosis and hemorrhage in number of studies<sup>63,64</sup> in patients with platelet counts of greater than  $5 \times 10^5$  cells/µL. The majority of these patients had reactive thrombocytosis, secondary to disseminated cancer (36%), infection (8 to 21%), or rebound from hemorrhage (19%). Only 6% of the thrombocytosis were due to myeloproliferative disorders. When patients with counts of over 10<sup>6</sup> platelets/µL were analyzed, 82% were due to reactive thrombocytosis. These were secondary to infection (31%), trauma (14%), malignant diseases (14%), and splenectomy (19%), while 14% were due to myeloproliferative disorders. Among the latter group, chronic myeloid leukemia constituted 42%, essential thrombocythemia 29%, and polycythemia vera 13%. Clinically, the thromboembolic complications were much more frequently seen in the myeloproliferative disorders than in reactive thrombocytosis. This is presumably due to the transient nature of the latter condition. Platelets in essential thrombcythemia and in polycythemia vera have been found to have a number of abnormal characteristics that increase the thrombotic risks.65 These include increased inhibitory activity against plasminogen activators and plasmin,66 abnormal aggregation,67 increased thromboxane generation, and increased thrombin generation due to the abnormal presence of phosphatidylserine on the surface of platelet membrane.68 On the other hand, there are also abnormalities that render the patients more liable to bleeding complications. These include decreased intracellular adenine nucleotides,69 decreased prothrombinase expression,<sup>70</sup> and deficient receptor function from decreased membrane glycoproteins.71

Thrombosis is more common in arteries than in veins, and the incidence is not directly correlated to the platelet count.72 The arterial thrombosis occurs in cerebral, retinal, cardiac, mesenteric, and peripheral arteries, resulting in the respective manifestations of transient ischemic attack (TIA), stroke, amaurosis fugax and blindness, myocardial infarction, bowel infarction and peripheral limb ischemia.73 Microcirculatory impairment may also give rise to the syndrome of erythromelagia, which consists of erythematous, painful and burning skin manifestations in the extremities and acrocyanosis.74 The risk of thrombosis is increased with comorbid conditions such as hypertension, obesity, hyperlipidemia, and diabetes and with cigarette smoking. Despite earlier reports of low thrombotic risks in younger patients,<sup>75,76</sup> it is now clear that the incidence of thrombosis or hemorrhage is just as high in the younger age group,77-79 and that treatment is indicated for all ages.

Treatment consists of antiplatelet agents in the form of aspirin, ticlopidine, or clopidogrel. The symptoms of erythromelagia are often dramatically relieved after several days of aspirin therapy.

Suppression of thrombocytosis can be accomplished by chemotherapy with hydroxyurea or analegride. Although there is a leukemogenic potential in long-term hydroxyurea therapy, it is unclear how significant this actually proves to be. Analegride, on the other hand, is a quinazolin derivative that interferes with megakaryocyte maturation<sup>80</sup> and does not carry the risk of leukemogenesis.<sup>81</sup> However, it can cause certain side effects related to fluid retention.

#### PLASMA COMPONENTS OF BLOOD

The hyperviscosity syndromes can also be due to abnormalities of the plasma components of blood. The majority of these abnormalities are the presence of ab-

TABLE	2. H	yperv	iscosi	ty Sy	/ndrome	•
with	Abno	ormal I	Plasma	a Pro	oteins	

Multiple myeloma Waldenström's macroglobulinemia Cryoglobulinemia Pyroglobulinemia Dysfibrinogenemia

normal immunoglobulins in blood in plasma cell dyscrasias (Table 2).

#### **Multiple Myeloma**

The paraproteins in multiple myeloma cause rheologic abnormalities by several mechanisms. These proteins increase red cell aggregation, seen in red cells shown in peripheral blood smears and are manifested in the peripherial blood as rouleaux formations (red cells stacked up in rows). The internal viscosity of the red cell is also increased with reduced red cell deformability.82 The increase in plasma viscosity is dependent on the quantity and quality of the paraproteins. The latter is determined by the molecular size and shape of the paraprotein. Thus, hyperviscosity syndrome is most common in Waldenström's macroglobulinemia, followed by IgA myeloma because of the frequent polymerization of the IgA paraprotein.83-85 In the case of IgM, the serum viscosity is proportional to the plasma concentration of this paraprotein up to 3 g/dL, then increases sharply at higher levels. In IgG myeloma with hyperviscosity syndrome, the paraprotein is usually that of IgG-3 class, as IgG-3 tends to form aggregates and complexes. The syndrome in IgG-3 myeloma is seen at a plasma level of 4 to 5 g/dL of this paraprotein, but in other forms of IgG, hyperviscosity is only encountered at levels above 15 g/dL. In IgA myeloma, the syndrome is seen at 6 to 7 g/dL with the polymeric form of IgA, whereas with the monomeric form the level has to reach 10 to 11 g/dL for symptoms to occur. Hyperviscosity is also seen in kappa light chain disease, again due to the formation of the light chain aggregates.

The degree of hyperviscosity is assessed by measurements of the serum viscosity, usually employing a capillary tube viscometer. The unit of viscosity is expressed as units of relative viscosity (relative to water of 1.0). The relative viscosity of normal serum is 1.4 to 1.8, while that of symptomatic patients is generally above 5. A more accurate rheologic analysis requires instruments that take into account the factor of shear rates, by placing the test liquid (whole blood, plasma, or serum) between two rotating surfaces. The viscosity at lower shear rates ranging from 0.18 to  $1.5 \text{ s}^{-1}$  can be measured. These results give a much better correlation with the clinical symptoms.

In addition to increased viscosity, hemostatic function is impaired at high paraprotein levels. Global tests of hemostasis such as bleeding time, prothrombin time, and activated partial thromboplastin time are often prolonged. The interaction between the paraproteins and the various clotting factors, including fibrinogen, factors V, VII, and VIII has been well studied. The paraproteins cause impaired reaction of these clotting factors at different stages of the cascade, resulting in the prolonged clotting times. Platelet function (adhesion and aggregation) is also impaired resulting in the lengthening of the bleeding time. These abnormalities are dependent on the level of the paraproteins and are reversible following therapy.

The clinical manifestations in this group of hyperviscosity syndrome are in many aspects similar to those seen in polycythemia vera. They are the results of vascular occlusion involving blood vessels of varying sizes and may affect any part of the body. In addition, there are also manifestations due to impaired hemostasis. Neurologic and visual symptoms are most frequent. Central nervous system manifestations include headache, dizziness, and impaired mentation up to coma. The impairment of retinal vein blood flow results in a characteristic fundal appearance of a "link-sausage" appearance, consisting of alternating venous segments with dilatation and constriction. This is associated with retinal hemorrhages and exudates. Peripheral neuropathy and myopathy due to occlusion of the neural vessels may also occur, often as the presenting symptoms.

The effectiveness of therapeutic apheresis of the paraproteins depends on the distribution of the proteins, whether they are intravascular or extravascular. The major fraction of macroglobulins and the larger aggregates of the other paraproteins are present in the intravascular space and can thus be effectively removed by plasma exchange. A plasmapheresis apparatus employing continuous flow removal is better tolerated by the patient and is most widely used. With the exchange of one plasma volume (39 to 44 mL per kg body weight), 65% of macroglobulin can be removed. If the IgM level before plasma exchange is high, the serum viscosity can be effectively reduced by 50% or more by just one exchange. However, the frequency of plasmapheresis is best guided by repeated serum viscosity measurements. This procedure is generally well tolerated with an incidence of complications of around 12 to 40%, usually of minor nature.86 These are commonly due to citrate toxicity manifesting as paresthesia, muscle twitching, and cramps or tetany, and can be readily corrected by calcium administration. Plasma exchange may also remove a substantial number of platelets, erythropoietin, and certain antibiotics.

Cryofibrinogenemia was first described with multiple myeloma<sup>87</sup> in a patient with cold-precipitable proteins in the plasma but not in serum. Since then, cryofibrinogen has been found in patients with leukemia and metastatic carcinomas, often associated with thrombophlebitis migrans. Cryofibrinogen is a complex of fibrinogen, cold-insoluble globulin, fibronectin, and precipitates at 4°C, and often at room temperatures. Occlusion of vessels may occur in the cooler parts of the body such as the skin in the extremities, the digits, the ears, and tip of the nose, as a result of cold precipitation of the cryofibrinogen. These occlusive lesions manifest as cutaneous ulcerations, "purpura," and livedo reticularis.<sup>88</sup>

Cryoglobulins are cold-precipitable proteins in serum.<sup>89</sup> Thus, at cooler parts of the body, they may pose the problem of impaired circulation as in cryofibrinogenemia. The incidence has changed dramatically in the last decade from being a complication seen in plasma cell dyscrasia to one that is associated with hepatitis C. They are classified according to their composition. Type I cryoglobulins are monoclonal immunoglobulins with IgM being the most common, followed by IgG and IgA. They may also be composed of light chains alone (Bence-Jones protein). They are seen in myeloma and Waldenström's macroglobulinemia, chronic lymphocytic leukemia, non-Hodgkins lymphoma, and benign monoclonal gammopathy, the latter also known as essential monoclonal cryoglobulinemia. Type II cryoglobulins, also known as mixed monoclonal cryoglobulins, are those with IgG complexes with an IgM, IgA or another IgG molecule. This form of cryoglobulin is found in infections, primarily hepatitis C, autoimmune disorders, and lymphoproliferative disorders. Type III cryoglobulins, also known as mixed polyclonal cryoglobulins, are complexes of one or more classes of polyclonal immunoglobulins and sometimes include complement and viral and other antigens. This group of cryoglobulins is associated with infections, most prominent being hepatitis C. It is seen in a variety of diseases in which a polyclonal immunoglobulin response occurs, such as in infections, parasitic infestation, and autoimmune disorders. The viral diseases include hepatitis A, hepatitis B, hepatitis C, infectious mononucleosis, and cytomegalovirus infection. Other associated infections include leprosy, syphilis, and lymphogranuloma venereum. Parasitic infestations include leishmaniasis, toxoplasmosis, and schistosomiasis. Autoimmune disorders include systemic lupus erythematosus, rheumatoid arthritis, and pemphigus. The manifestations are similar to those seen in cryofibrinogenemia.

Pyroglobulins are precipitable from serum at 56°C and redissolve when the serum is cooled. They are not found as commonly as cryoglobulins. Again, they can be found in multiple myeloma and Waldenström's macro-globulinemia.

# HYPERVISCOSITY IN SOLID TUMORS

In recent years there were several reports showing that a hyperviscosity syndrome is present in patients with a wide variety of malignant tumors, whose clinical

course was complicated by thromboembolic events, usually deep vein thrombosis. Rheologic studies revealed that the abnormality was independent of the hematocrit.90 These malignant tumors include carcinoma of breast,<sup>91,92</sup> ovary,<sup>90</sup> lung,<sup>93</sup> head and neck,<sup>94</sup> and in melanoma.95,96 The factors responsible for the rheologic abnormality include increased fibrinogen levels, presence of fibrinogen degradation products, host inflammatory acute-phase proteins,97 and perhaps other unknown causes that have yet to be discovered. In addition to causing increased plasma viscosity, all of these factors also lead to an increase in red cell aggregation.94 The correlation of the increased viscosity with the stage of the cancer and the prognosis has not been established.<sup>90</sup> However, the clinician should be aware of this abnormality, as this is associated with increased risk for thromboembolic complications.

# SUMMARY AND CONCLUSION

The wide variety of pathologic conditions that can be complicated by the occurrence of the hyperviscosity syndrome share one common pathophysiologic feature. They all have abnormal rheologic features, leading to impaired circulation of blood. A thorough understanding of the rheologic abnormalities will help explain the clinical manifestations, especially in the location of the vascular occlusion. Identification of the responsible pathologic component of blood is needed to enable its prompt removal from blood and direct the therapeutic approach to control its production.

## REFERENCES

- Ross JM, Alevriadon BR, McIntire LV. In: Loscalzo J. Schafer A, ed. Thrombosis and Hemorrhage, 2nd ed. Philadelphia PA: Williams & Wilkins; 1998:405–421
- Lowe GDO. Blood rheology in venous thrombosis. Clin Hemorheol 1984;4:571–573
- Dintenfass L. Haemorrhage of cancer metastasis: An example of multiple myeloma, survival times and abnormality of blood viscosity factors. Clin Hemorheol 1982;2:259–271
- Chien S. Hemorheology in disease, pathophysiological implications. Clin Rheol 1981;1:319–322
- Jen CJ, McIntire LV. Characteristics of sheer-induced aggregation in whole blood. J Lab Clin Med 1984;103:115–124
- Wagner CT, Kroll MH, Chow TW, et al. Epinephrine and sheer stress synergistically induced platelet aggregation via a mechanism that partially bypasses v.Wf-GPIb interactions. Biorheology 1996;33:209–229
- Bell WR. Defibrinogenation with Arvin<sup>®</sup>. In: Brinkhouse KM, Hinnmon S, eds. Thrombosis: Mechanism and Control. Stuttgart: FA Schattauer Verlag; 1973:371–375
- Chien S. Biophysical behavior of red cells in suspensions. In: McN SD, ed. Red Blood Cell, Vol. 3, New York: Academic Press; 1975:1031-1133
- Chien S, Usami S, Bertles JF. Abnormal rheology of oxygenated blood in sickle cell anemia. J Clin Invest 1970;49:623–634

- Kroll MH, Hellums JD, McIntire LV, Shafer AI, Moake JL. Platelets and shear stress. Blood 1996;88:1525–1541
- Sharefkins JB, Diamond SL, Eskin SG, Dieffenbach CW. Fluid flow decreases preproendothelin mRNA levels and suppresses endothelin-1 peptide release in cultured endothelial cells. J Vasc Surg 1991;14:1-9
- Iba T, Shin T, Sonoda T, Rosales O, Sumpio BE. Stimulation of endothelial secretion of tissue-type plasminogen activator by repetitive stretch. J Surg Res 1991;50:457–460
- Diamond SO, Eskin SG, McIntire LV. Fluid flow stimulates tissue plasminogen activator secretion by cultured human endothelial cells. Science 1989;243:1483-1485
- Grabowski EF, Jaffe EA, Weksler BB. Prostacyclin production by cultured endothelial cell monolayers exposed to step increases in shear stress. J Lab Clin Med 1985;105:36–34
- Carosi JA, McIntire LV. Modulation of secretion of vasoactive materials from human and bovine endothelial cells by cyclic strain. Biotechnol Bioengin 1994;43:615–621
- Chien S, Li S, Shyy YJ. Effects of mechanical forces on signal transduction and gene expression in endothelial cells. Hypertension 1998;31:162–169
- Lin MC, Almus-Jacobs F, Chen HH, et al. Shear stress induction of the tissue factor gene. J Clin Invest 1997;99:734–744
- Moake JL, Turner NA, Stathopoulos NA, Nolasco L, Hellums JD. Shear induced platelet aggregation can be mediated by vWF releases from platelets, as well as by exogenous large or unusually large vWF multimers, requires adenosine diphosphate, and is resistant to aspirin. Blood 1988;71:1366–1374
- Wasserman LR, Balcerzak SP, Berk PD, et al. Influence of therapy on causes of death due to polycythemia vera. Trans Ass Am Physicians 1981;94:30-38
- Gruppo Italiano Study Policitemia. Polycythemia vera: The natural history of 1213 patients followed for 20 years. Ann Intern Med 1995;123:656-664
- 21. Berlin NI. Polycythemia vera. Semin Hematol 1997;34:1-78
- Fruchtman SM, Berk PD. Polycythemia vera and angiogenic myeloid metaplasia. In: Handin RI, Lux SE, Stossel TP, eds. Blood, Principles and Practices of Hematology. Philadelphia PA: Lippincott; 1995:415–438
- Means RT Jr. Polycythemia vera. In: Lee RG, Foerster J, Lukens J, Paraskevas F, Greer JP, Rodgers GM, eds. Wintrobe's Clinical Hematology. Baltimore, MD: Williams and Wilkins; 1998:2374–2389
- Wells RE Jr., Merrill EW. The variability of blood viscosity. Am J Med 1961;31:505–509
- Dintenfass L. Haemorheology and the microcirculation: Role of the inversion phenomenon. In: Garlick D, ed. Progress in Microcirculation Research. Sidney: Committee in postgraduate medical education, University of New South Wales; 1980: 100-104
- Dintenfass L, Sommer T. On aggregation of red cells in Waldenström's macroglobulinemia and multiple myeloma. Microvasc Res 1975;9:279–286
- Dintenfass L. Significance of ABO blood-groups on aggregation of red cells in patients with diabetes mellitus. Role of fibrinogen and serum proteins. Microvasc Res 1974;7:326–341
- Dintenfass L. Haemorheology of cancer metastases: An example of malignant melanoma, survival times and abnormality of blood viscosity factors. Clin Hemorheol 1982;2:259–271
- Kwaan HC, Levin A, Sakurai S, et al. Digital ischemia and gangrene due to red blood cell aggregation induced by acquired dysfibrinogenemia. J Vas Surg 1997;26:1061–1068
- Dintenfass L. Aggregation of red cells in disease: Some deductions and speculations based on the results of "ARC" experiments on the space shuttle "Discovery" STS 51. Clin. Biorheol 1988;25:65-76
- Carlos TM, Harlan JM. Leukocyte-endothelial adhesion molecules. Blood 1994;84:2068–2101

- Joseph-Silverstein J, Silverstein RL. Cell adhesion molecules: An overview. Cancer Invest 1998;16:176–182
- Bevilacqua MP. Endothelial-leukocyte adhesion molecules. Annu Rev Immunol 1993;11:767–804
- Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: The multistep paradigm. Cell 1994;76: 301-314
- Butcher EC. Leukocyte-endothelial cell recognition: Three (or more) steps to specificity and diversity. Cell 1991;67:1033– 1036
- Newham P, Humphries MJ. Integrin adhesion receptors: Structure, function and implications for biomedicine. Mol Med Today 1996;2:304–313
- Roth GT. Developing relations: Arterial platelet adhesion, glycoprotein Ib, and leucine-rich glycoproteins. Blood 1991;77:5– 19
- Lawrence MBO, McIntire LV, Smith CW, Eskin SG. Effect of venous sheer stress on CD18 mediated neutrophil adhesion to cultured endothelium. Blood 1990;75:227-237
- Smith CW, Kishimoto TK, Abbassio O, et al. Chemotatic factors regulate LEC-CAM 1 dependant neutrophil adhesion to cytokine stimulated endothelial cells in vitro. J Clin Invest 1991;87:609-618
- Grunwald GB. The structural and functional analysis of cadherin calcium-dependant cell adhesion molecules. Curr Opin Cell Biol 1993;5:797–805
- Takeichi M. The cadherins: Cell-cell adhesion molecules controlling animal morphogenesis. Development 1998;102:639– 655
- Su LK, Vogelstein B, Kinzler KW. Association of the APC tumor suppressor protein with cadherins. Science 1993;262:1734– 1737
- Bevilacqua MP, Pober JS, Mendrick DC, et al. Identification of an inducible endothelial-leukocyte adhesion molecule. Proc Natl Acad Sci USA 1987;84:9238–9242
- Gallatin WM, Weissman II, Butcher EC. A cell surface molecule involved in organ-specific homing of lymphocytes. Nature 1983;304:30–34
- Hsu-Lin SC, Berman CL, Furie BC, et al. A platelet membrane protein expressed during platelet activation. J Biol Chem 1984;259:9121-9126
- Albelda SM. Role of cell adhesion molecules in tumor progression and metastasis In: Wegner CD, ed. The Handbook of Immunopharmacology: Adhesion Molecules. New York: Academic Press, Harcourt Brace; 1994:71–88
- Sass PM. The involvment of selectins in cell adhesion, tumor progression, and metastasis. Cancer Invest 1998;16:322–328
- Sanders RJ, Mainero F, Giancotti FG. The role of integrins in tumorogenesis and metastasis. Cancer Invest 1998;16:329–344
- Jain RK, Koenig GC, Dellian M, et al. Leukocyte-endothelial adhesion and angiogenesis in tumors. Cancer Met Rev 1996;15: 195-204
- Aul C, Gatterman N, Germing U, et al. Fatal hyperleukocytic syndrome in a patient with chronic myelomonocytic leukemia. Leukemia Res 1997;21:249–253
- Lawrence MB, Springer TA. Leukocytes roll on a selection at physiological flow rates: Distinction from and prerequisite for adhesion through integrins. Cell 1991;65:859–873
- Ruco LP, Pittiglio M, Dejana E, Baroni CD. Vascular activation in the histopathogenesis of Hodgkin's disease: Potential role of endothelial tissue factor in intravascular thrombosis and necrosis. J Pathol 1993;171:131–136
- Lichtman MA. Rheology of leukocytes, leukocyte suspensions, and blood in leukemia. Possible relationship to clinical manifestations. J Clin Invest 1973;52:350–358
- McGee LC Jr, Collins RD. Intravascular leukocyte thrombi and aggregates as a cause of morbidity and mortality in leukemia. Medicine 1974;53:463–478

- Nowacki P, Fryze C, Zdziarska B, et al. Central nervous system leukostasis in patients with leukemias and lymphomas. Folia Neuropathol 1995;33:59–65
- Fritz RD, Forkner CD Jr, Freirich EJ. The association of fatal intracranial hemorrhage and "blastic crisis" in patients with acute leukemia. N Engl J Med 1959;294:245–248
- Vernant JP, Brun B, Mannoni P, Dreyfus B. Respiratory distress of hyperleukocytic granulocytic leukemias. Cancer 1979;44: 264–268
- Myers TJ, Cole SR, Klatsky AU, Hild DH. Respiratory failure due to pulmonary leukostasis following chemotherapy of acute nonlymphocytic leukemia. Cancer 1983;51:1808–1813
- Cuttner J, Holland JF, Norton L, et al. Therapeutic leukapheresis for hyperleukocytosis in acute myelocytic leukemia. Med Ped Oncol 1983;11:76–78
- Eguiguren JM, Schell MJ, Crist WM, Kunkel K, Rivera GK. Complications and outcome in childhood acute lymphoblastic leukemia with hyperleukocytosis. Blood 1992;79:871–875
- Berg J, Vincent PC, Gunz FW. Extreme leukocytosis and prognosis of newly diagnosed patients with acute nonlymphocytic leukemia. Med J Aust 1979;1:480–482
- Sharma K, Rao S, Bhat SV. Effect of hydroxyurea on blood viscosity in chronic myelogenous leukemia with hyperleukocytosis. Physiol Chem Phys Med NMR 1991;23:261–268
- Davis WM, Ross AO. Thrombocytosis and thrombocythemia: The laboratory and clinical significance of an elevated platelet count. Am J Clin Pathol 1973;59:243–247
- Sarthosh-Kumar CR, Yohannan MD, Higgy KE, Al-Mashhadani SA. Thrombocytosis in adults: Analysis of 777 patients. J Intern Med 1991;229:493–495
- Schafer AI. Bleeding and thrombosis in the myeloproliferative disorders. Blood 1984;64:1–12
- Kwaan HC, Suwanwela N. Inhibition of fibrinolysis in platelets in polycythemia vera and thrombocytosis. Br J Haematol 1971;21:313–322
- Baker RI, Manoharan A. Platelet function in myeloproliferative disorders: Characterization and sequential studies show multiple platelet abnormalities, and change with time. Eur J Haematol 1988;40:267–272
- Lee LH, Baglin T. Altered platelet phospholipid-dependant thrombin generation in thrombocytopenia and thrombosis. Br J Haematol 1995;89:131-136
- Leoncini G, Maresca M, Buzzi E et al. Platelets of patients affected with essential thrombocythemia are abnormal in plasma membrane and adenine nucleotide content. Eur J Haematol 1990;44:115–119
- Crook M, Machin SJ, Crawford N. Basal and induced prothrombinase expression in platelets from patients with essential thrombocythemia. Br J Haematol 1990;76:256–259
- Mazzucato M, DeMarco L, DeAngelis V, et al. Platelet membrane abnormalities in myeloproliferative disorders: Decrease in glycoproteins Ib and IIb/IIIa complex is associated with deficient receptor function. Br J Haematol 1989;73:369–374
- Barbui T, Cortelazzo S, Viero P, et al. Thrombohaemorrhagic complications in 101 cases of myeloproliferative disorders: Relationship to platelet number and function. Eur J Cancer Clin Oncol 1983;19:1593-1599
- Hussain S, Schwartz JM, Friedman SH, Chua SN. Arterial thrombosis in essential thrombocythemia. Am Heart J 1978;96: 31-36
- Michiels JJ, VanGenderen PJJ, VanVliet HHDM. Erythomelalgia and arterial thrombophilia in thrombocythemia. Ann NY Acad Sci 1994;714:318–321
- Hoagland HC, Silverstein MN. Primary thrombocythemia in the young patient. Mayo Clin Proc 1978;53:578–580
- McIntyre KJ, Hoagland HC, Silverstein MN, Pettit RM. Essential thrombocythemia in young adults. Mayo Clin Proc 1991;66: 149–154

- Mitus AJ, Barbui T, Shulman LN et al. Hemostatic complications in young patients with essential thrombocytosis. Am J Med 1990;88;371–375
- Watson KV, Key N. Vascular complications of essential thrombocythemia. A link to cardiovascular risk factors. Br J Haematol 1993;83:198-203
- Millard FE, Hunter CS, Anderson M, et al. Clinical manifestations of essential thrombocythemia in young adults. Am J Hematol 1990;33:27-31
- Mazur EM, Rosmarin AG, Sohl PA, Newton JL, Narendran A. Analysis of the mechanism of anagrelide-induced thrombocytopenia in humans. Blood 1992;79:1931–1937
- Petitt RM, Silverstein MN, Petrone ME. Anagrelide for control of thrombocythemia in polycythemia and other myeloproliferative disorders. Semin Hematol 1997;34:51-54
- McGrath MA, Penny R. Paraproteinemia, blood viscosity and clinical manifestations. J Clin Invest 1976;58:1155–1162
- Farhangi M, Osserman EF. Biology, clinical patterns, and treatment of multiple myeloma and related plasma cell dyscrasias. In: Twomey JJ, Good RA, eds. The Immunopathology of Lymphoreticular Neoplasms, Comprehensive Immunology. New York: Plenum Medical Book Co; 1978:641–719
- Block KJ, Maki OG. Hyperviscosity syndromes associated with immunoglobulin abnormalities. Semin Hematol 1973;10:113–124
- Crawford J, Cox EB, Cohen HJ. Evaluation of hyperviscosity in monoclonal gammopathies. Am J Med 1985;79:13-22
- Sutton DMC, Nair RC, Rock G, et al. Complications of plasma exchange. Canadian Apheresis Study Group. Transfusion 1989;26:124–127
- Wintrobe MM, Buehl MV. Hyperproteinemia associated with multiple myeloma. Bull John Hopkins Hosp 1933;52:156–165

- Ireland TA, Werner DA, Reitschel RL, et al. Cutaneous lesions in cryofibrogenemia. J Pediatr 1984;105:67–69
- Foerster J. Cryoglobulins and cryoglobulinemia. In: Lee GR, Foerster J, Lukens J, Paraskevas F, Greer JP, Rodgers GM, eds. Wintrobe's Clinical Hematology Baltimore, Williams & Wilkins; 1998:2725-2737
- von Templehoff G-F, Heilmann L, Hommel G, et al. Hyperviscosity syndrome in patients with ovarian carcinoma. Cancer 1998;82:1104–1111
- Miller B, Heilman L. Hemorheologic variables in breast cancer patients at the time of diagnosis and during treatment. Cancer 1988;62:350-354
- von Templehoff G-F, Heilman L. Thrombosis and hemorheology in patients with breast cancer and adjuvant chemotherapy. Clin Hemorheol 1995;15:311–323
- Chen H, Yunpeng W, Shaoxi C, Mian L. A research on rheological properties of erythrocyte of lung cancer patients. Clin Hemorheol 1996;16:135-141
- Khan MM, Puniyani RR, Huilol NG, Hussain MA, Ranade GG. Hemorheological profiles in cancer patients. Clin Hemorheol 1995;15:37-44
- Dintenfass L. Hemorheology of cancer metastasis: An example of malignant melanoma, survival times and abnormality of blood viscosity factors. Clin Hemorheol 1982;2:259–271
- Dintenfass L. Criteria and values in diagnosis and treatment and prevention of metastasis in neoplastic diseases. J Clin Pathol 1987;88:596-602
- Weng X, Cloutier G, Beaulieu R, Roederer GO. Influence of acute-phase proteins on erythrocyte aggregation. Am J Physiol 1996;271:H2346-H2352